

## Likely q's on SA Bio206 test

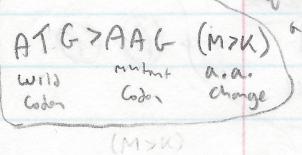
3.9-3.4

### ① Properties of genetic code:

- Non overlapping - Successive triplets read in order (each nt only part of one codon)
- Comma-free
- Degenerate - Some amino acids have multiple codons,
- triplet - 3nt code for 1 a.a.
- Unambiguous - each codon specifies only 1 particular a.a. or stop (start codon)
- Nearly universal - Only DNA is mitochondrial and some protozoans have different codes.

#### Genetic Code

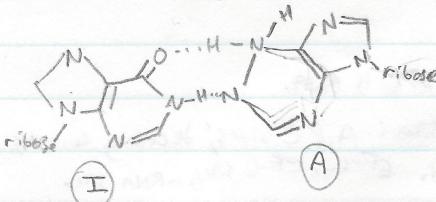
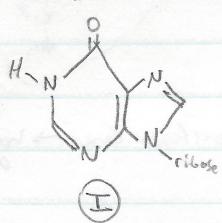
↳ translates nucleic acid sequence of mRNA into seqs of proteins.



### ② Degenerate code minimizes mutation effects?

- Codon {
- point mutation can cause a.a. substitution - try, Met → only 1 codon, since try = biggest, Met = start codon, so most deletions are substituted accidentally.
  - 1st a.a. → similar amino acids coded
  - 2nd a.a. → purine → pyrimidine → mostly polar a.a.'s if purine → pyrimidine then w/ get neg charge.
  - 3rd a.a. very degeneracy - even w/ change many still code for same amino acids - mutations often "silent"

### ③ Draw inosine, (draw non complementary base pairing?)



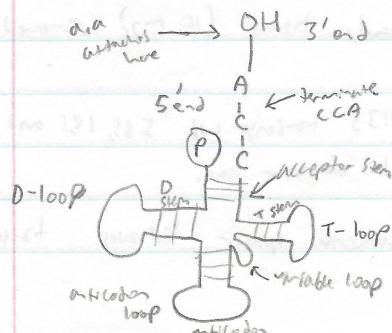
### ④ 3 types of RNA

3 rRNA  
for protein synthesis

- mRNA → codes for proteins
- rRNA → structures in ribosomes
- tRNA → carry a.a. from cytoplasm → ribosome bound to polypeptide.
- Small RNA → pre-mRNA splicing, rRNA processing, etc...

1st base in anticodon = wobble,  
2,3 = wobble (sometimes)

### ⑤ tRNA: draw it, describe it



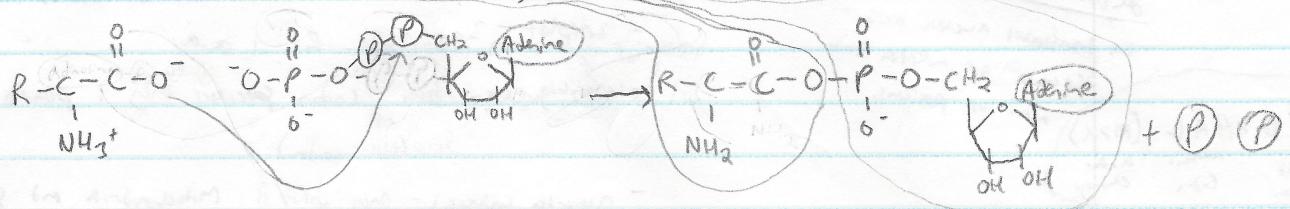
20 a.a.s → 61 codons → 31 tRNAs → 20 tRNAs (one per a.a.)  
 (isoreading tRNAs) ↑  
 3' start/stop → bind to multiple codons w/ wobble base pairing.  
 has modified bases in it.  
 ↓  
 UAG-amber  
 UAA-octre  
 UGA-opal  
 ↓  
 amino acid + tRNA

## ⑥ aaRS charging rxn...

↳ 2 classes... Class I → aminoacylates at the 2'-OH  
II " " 3'-OH

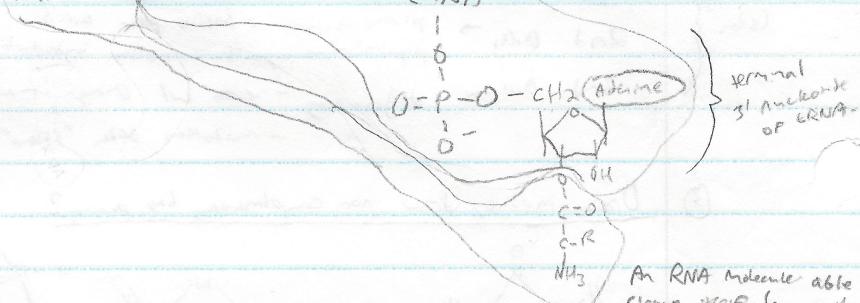
\* reversible, Anticodon is only link to mRNA Sequence.

2 steps



2 step rxn catalyzed by aminoacyl-tRNA Synthetase

(charging tRNA)



Ribozymes - 23S, 28S, 5rRNA

⑦ Ribosomes -  $\approx 70\%$  mass + 85% is RNA.

- 5 active sites: A, P, E sites; decoding centre; peptidyl transferase centre → have catalytic RNA protein at active sites.  
\* also EF-Tu, EF-G sites, mRNA sites.

### Prokaryote

- 70S ribosome

rRNA 7, 30S ribosome subunit

↳ 16S, 21 proteins

- 50S ribosome subunit

↳ 5S, 23S, 34 proteins

### Eukaryote

- 80S ribosome

- 540S ribosome subunit → matches tRNA to codons

↳ 18S, 33 proteins

A cavity b/w subunits fits 2 tRNAs.

- 60S ribosome subunit → Catalyzes peptide bond formation (23S, 28S)

↳ 5S, 5.8S, 28S, 49 proteins

## ⑧ tRNA, rRNA processing?

tRNA → RNAP III, 5', 3' ends removed, CCA added to 3' end, introns (if any) removed by mitochondrial endonucleases.

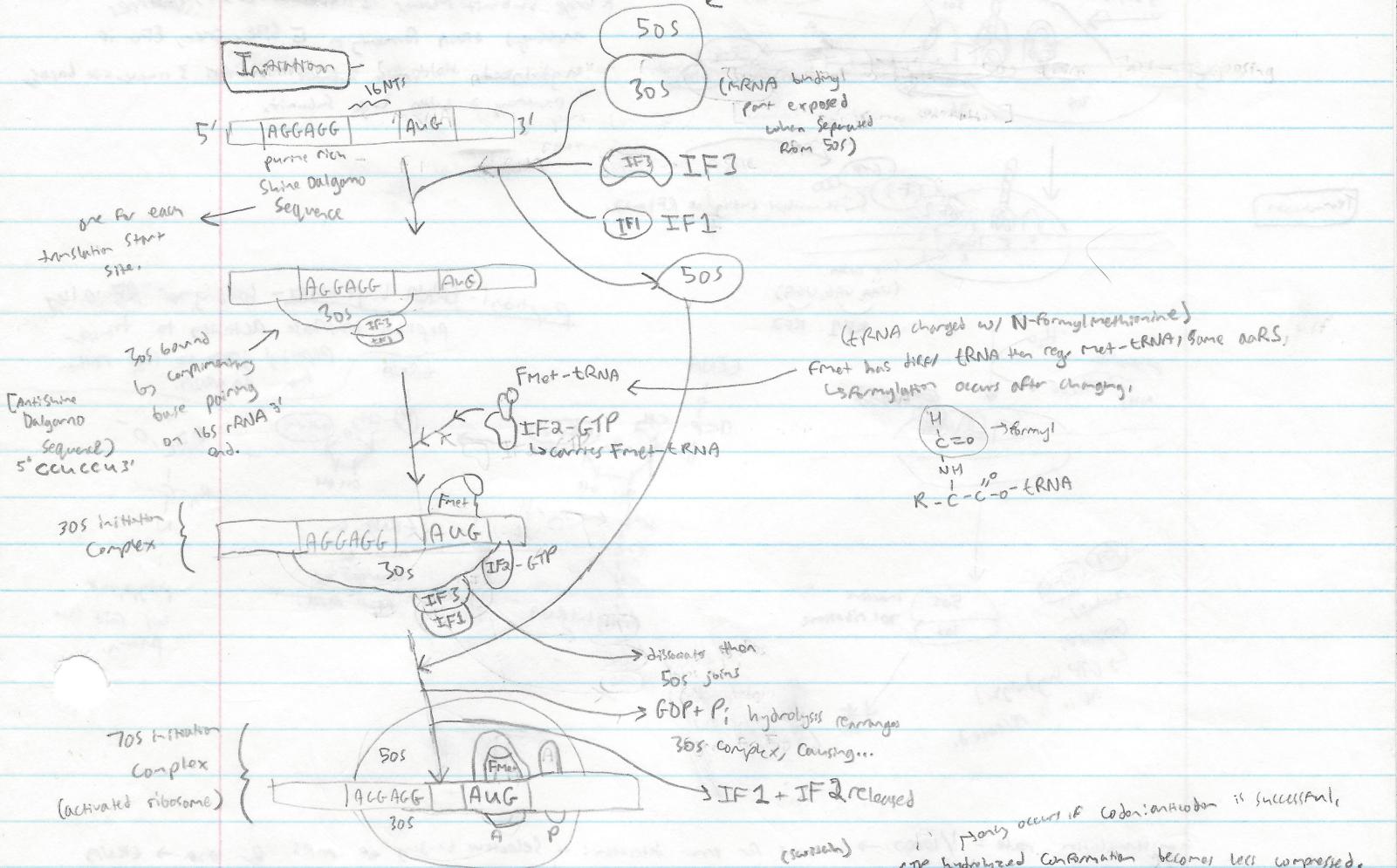
rRNA → rDNA genes in nucleolus, 2 genes: - hnRNA 43S transcript w/ 5.8S, 18S and 28S  
↳ RNAPI

- 5S coded on own gene.

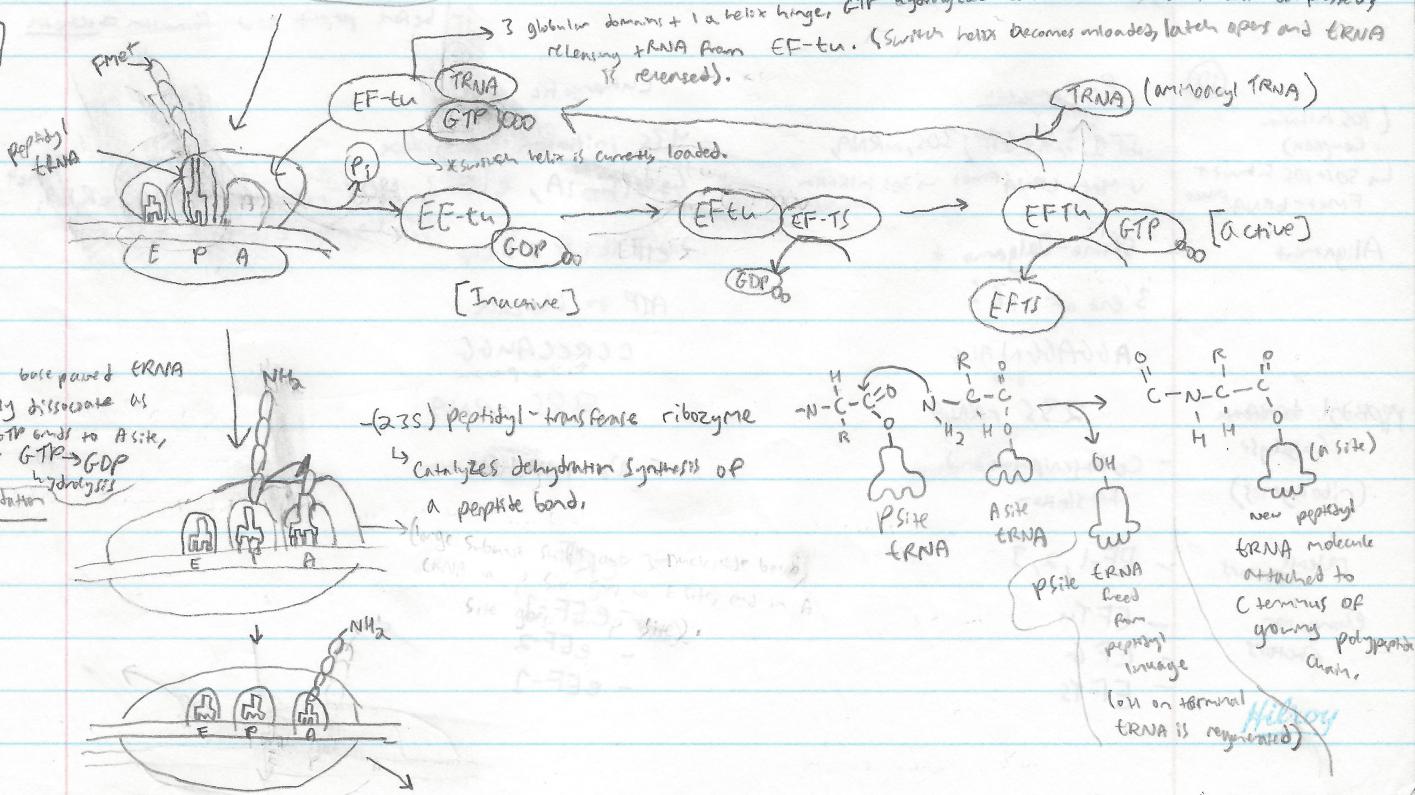
hnRNA 43S → 5', 3' ends removed, 18S cleaved,

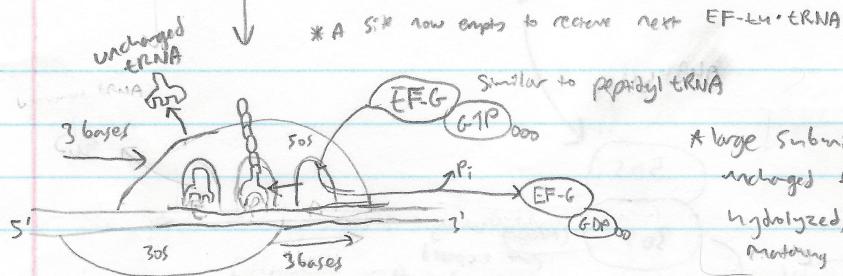
5.8S rearranges to 28S then loops. \* a nontranscribed spacer separates transcription units.

## ① Translation Steps



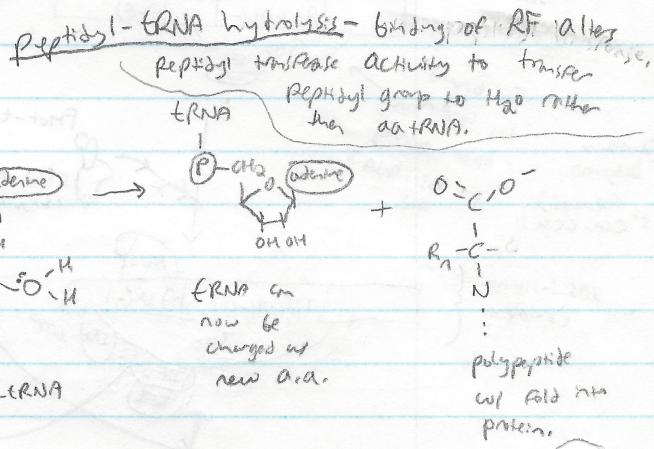
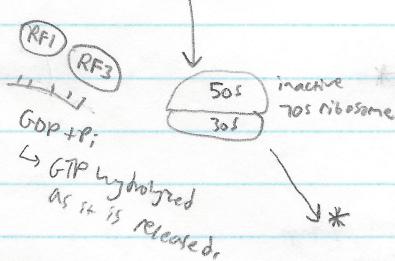
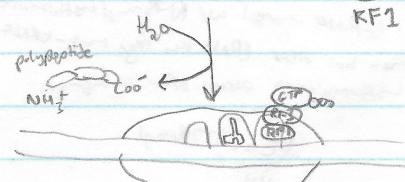
## Elongation





A large Subunit Moves 3 nucleotide bases, releases uncharged tRNA formerly in E Site. Once, EF-G is hydrolyzed, the small subunit shifts 3 nucleotide bases, moving it with large subunit.

Termination



- mistranslation rate = 1/10000 → 2 means for error detection:  
 - selective binding of aaRS for aa → tRNA  
 - before peptide bond formation = kinetic

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### Prokaryotic

(70S initiation complex)  
 L 50S+30S Subunit + FMet-tRNA<sub>FMet</sub>

- IF1, 3, 2 + GTP; 30S, mRNA, FMet-tRNA<sub>FMet</sub> → 70S initiation complex

Alignment

- Shine-Dalgarno + 3' end of 16S

(AGGAGGGN AUG  
8-15)

23S rRNA

### Eukaryotic

- 43S initiation complex

L eIF-1A, eIF-3, 40S ribosome, mRNA, tRNA<sub>Met</sub> + eIF-2 + GTP

- eIF4E + G + 5' cap

also: eIF4A, G, E.

eukaryotic  
↓ factor  
initiation

ATP to Kozak

CCRCCAUAG  
C is a purine

28S rRNA

- No cotranscription

- eRF

- eEF-1  
- eEF-2  
- eEF-3

release factors

- RF-1, 2, 3

elongation factors

- EFTu  
- EF-G  
- EFTs

⑩ - After translation - Chaperones may assist in folding

- post translational modifications: - Cleavage of initiator Met

- cleavage of Signal Sequences

- add of Co-factors

- glycosylation, acetylation, phosphorylation

- multiunit proteins assemble

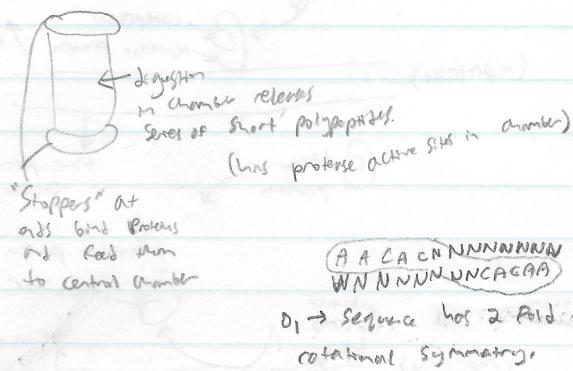
- transport to site of action

- tagged w/ ubiquitin attached to lysine side

chain of proteins.

lysines on proteins meant to be destroyed  
damaged w/ area exposed.

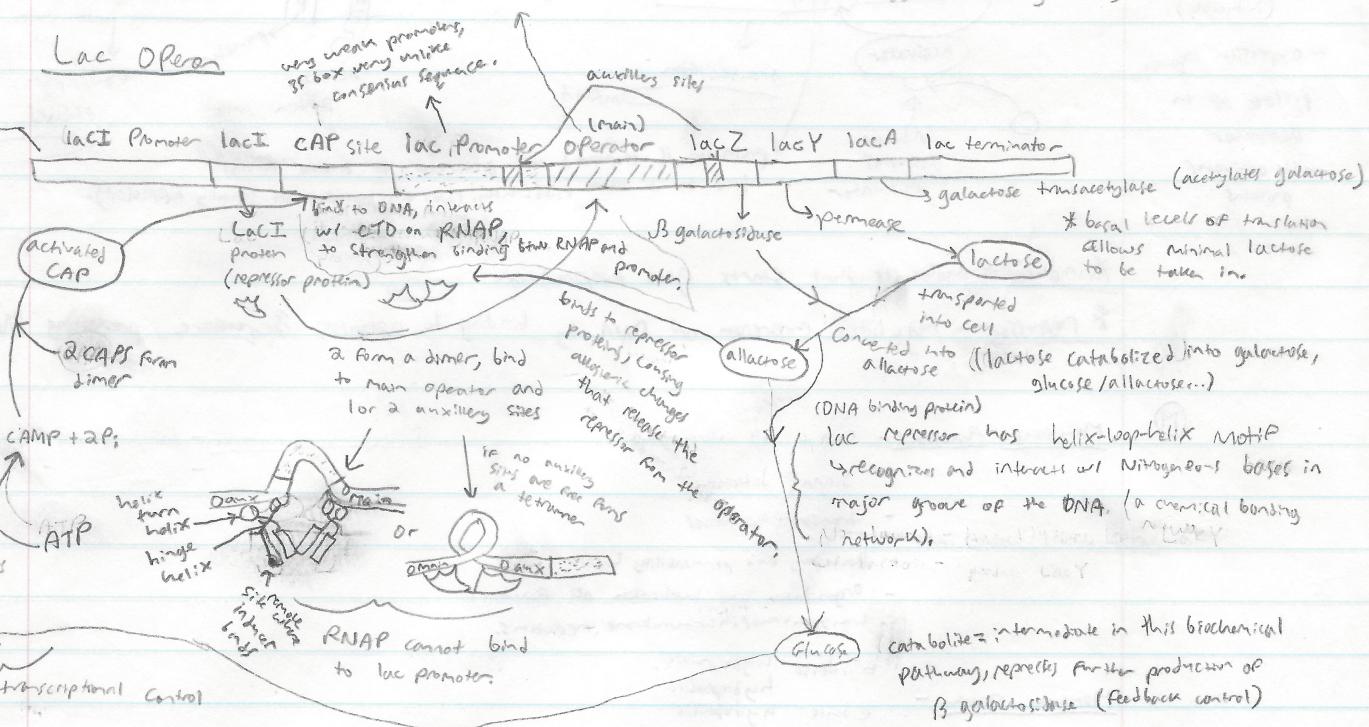
→ IP tagged then degraded by proteasomes.



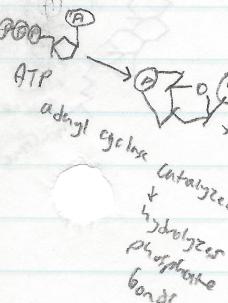
### Lac Operon

2 promoters:

$\sigma_70$ , or  $\sigma_S$



intracellular signal  
(↓ glucose ↑ cAMP)



### Attenuation

RNA formed interferes w/ transcription.  
- regulatory proteins can prevent b/w RNA w/ RNAP.

### Riboswitches

II (RNAP) → III (RNAP)  
R riboswitch binds to mRNA

Conformational changes, RNAP dissociates.

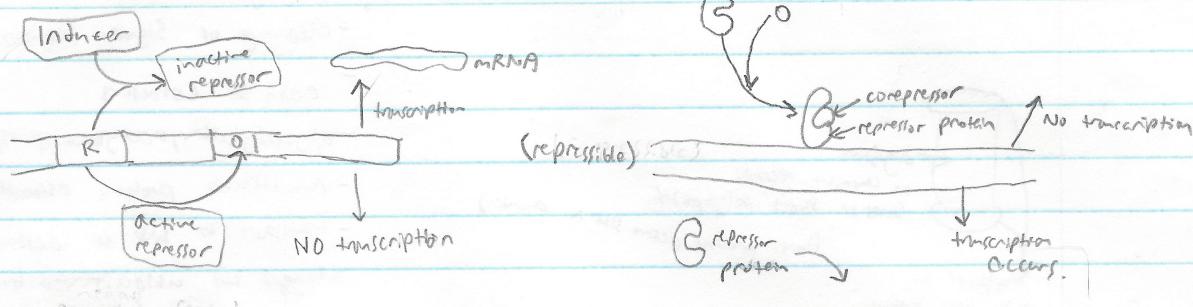
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## Positive vs. Negative Control

-ve Control

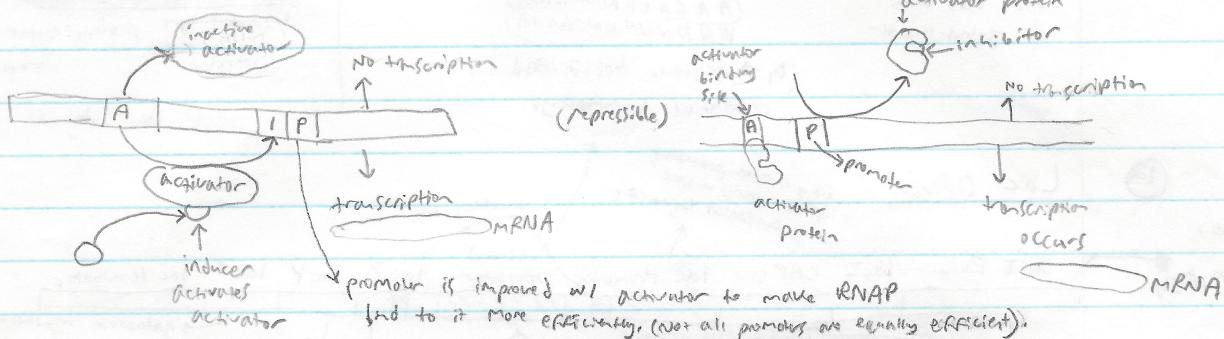
(inducible)

- expression normally blocked
- uses repressor proteins

Positive Control

(inducible)

- expression requires presence of M
- uses activator proteins

\*inducer - molecule that starts gene expression\*repressor - regulates expression of DNA by binding to operator sequence, preventing RNAP transcription

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## Membrane functions - Cell to cell interactions

- signal detection
- transport processes
- boundary and permeability barrier
- organization and localization of function
- transmembrane/in membrane reactions.

Membrane structure -

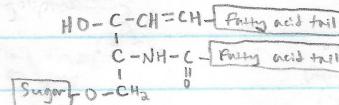
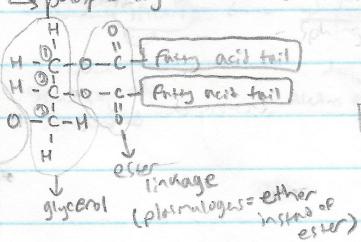
- composed of lipids and proteins 4:1 → 1:9 ratio.

- membrane lipids are asymmetrical (polar head/ nonpolar tail) so bilayers, bilayers are stabilized by hydrophobic interactions.

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## Lipids

↳ phosphatidyl... (alcohol) alcohol

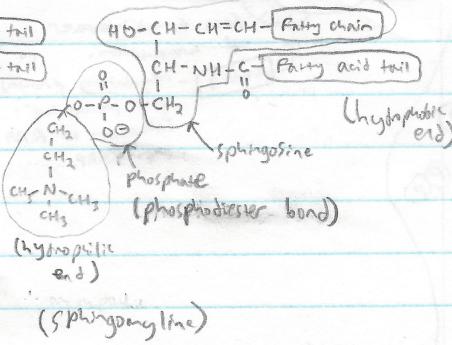


\* C2 usually longer and more double bonds

common alcohols → Choline, ethanolamine, serine → has net fee charge

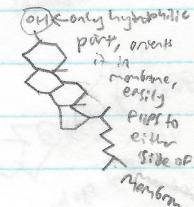
glycerol, inositol

↳ Sphingolipid



↳ short a choline head

↳ Sterol



properties - hydrophobic tail, hydrophilic head  $\rightarrow$  amphiphatic (polarity, directionality)

- spontaneously form vesicles, liposomes, bilayers  $\rightarrow$  thermodynamically favorable - minimize hydrophobic interactions w/ H<sub>2</sub>O.
- tails differ in saturation, length.
- membranes

fluidity depends on:  
- temp  $\rightarrow$  ↓ = less rotation and flexion motions  $\rightarrow$  fluidity  
- composition  $\leftrightarrow$  ↑ cholesterol  $\rightarrow$  ↓ fluidity  
- length of fatty acid  $\rightarrow$  ↑ = more hydrophobic interactions  $\rightarrow$  ↓ fluidity  
- saturation of fatty acids  $\rightarrow$  ↑ = less bends, more compact  $\rightarrow$  ↓ fluidity

### Lipid Rafts - specialized membrane

Microdomains compartmentalize cellular processes. ordered together but float freely in

the membrane bilayer.  
- Membrane synthesized in ER, inserted into cytosolic free, flippase catalyzes transfer of phospholipid molecule to outer face.

\* Scramblase flips phospholipid from cytosolic half of ER to ER lumen half of bilayer.  
 $\rightarrow$  req. longer to solubilize the membrane  
Protein  $\sim$  25 a.a domain (in bilayer)

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Proteins - Membrane proteins - 2 classes: integral  $\rightarrow$  transmembrane

Functions  $\rightarrow$  transport

$\rightarrow$  Anchors

$\rightarrow$  Receptors

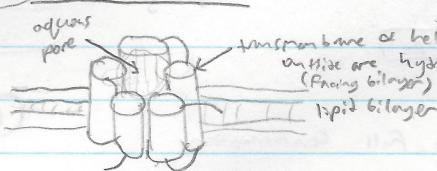
$\rightarrow$  Enzymes

myristoyl anchor  $\rightarrow$  amide

- GPI anchor - cytoskeleton anchor

using prenyl group  $\rightarrow$  palmitoyl anchor  $\rightarrow$  thioester

GPI anchors protein by covalently linking ester bond terminus of protein to glycolipid head group



hydrophobic a.a. side chain

hydrophobic a.a. backbone

lipid bilayer

### 16 strand β-barrel porin



- relatively non-selective, molecules < 600 daltons pass, larger ones do not

- side groups facing inward  $\rightarrow$  hydrophilic, outward  $\rightarrow$  hydrophobic.

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Cytoplasm - peripheral membrane proteins

$\rightarrow$  Filamentous proteins (intermediate, actin, microtubule) and accessory proteins

- reinforced w/ protein meshwork  $\rightarrow$  cytoskeleton (cell cortex)

- actomyosin

- coated w/ glycoproteins (proteins post-translationally modified to add glucose) + glycolipids.  
 $\hookrightarrow$  oligosaccharides exterior surfaces only!

- organelles - single membrane - ER, Golgi, lysosomes, endosomes, peroxisome

- double membrane - nucleus, mitochondria, chloroplast

$\hookrightarrow$  mutually incompatible chemical reactions must be able to take place simultaneously in some cells need segregation.

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Signal sequence for nucleus: Pro-Pro-Lys-Lys-Lys-Arg-Lys-al (string of two charged a.a.s)

Lys can have multiple, none

P P K K K R K V

Hilary

↳ usually removed at final destination

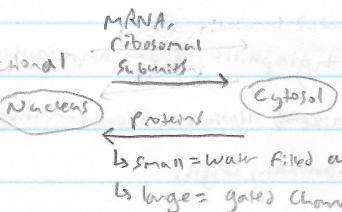
$\hookrightarrow$  stay in cytosol

(perinuclear space)

outer nuclear membrane continuous w/ ER,  $\rightarrow$  lumen in ER continuous w/ intermembrane lumen in nucleus

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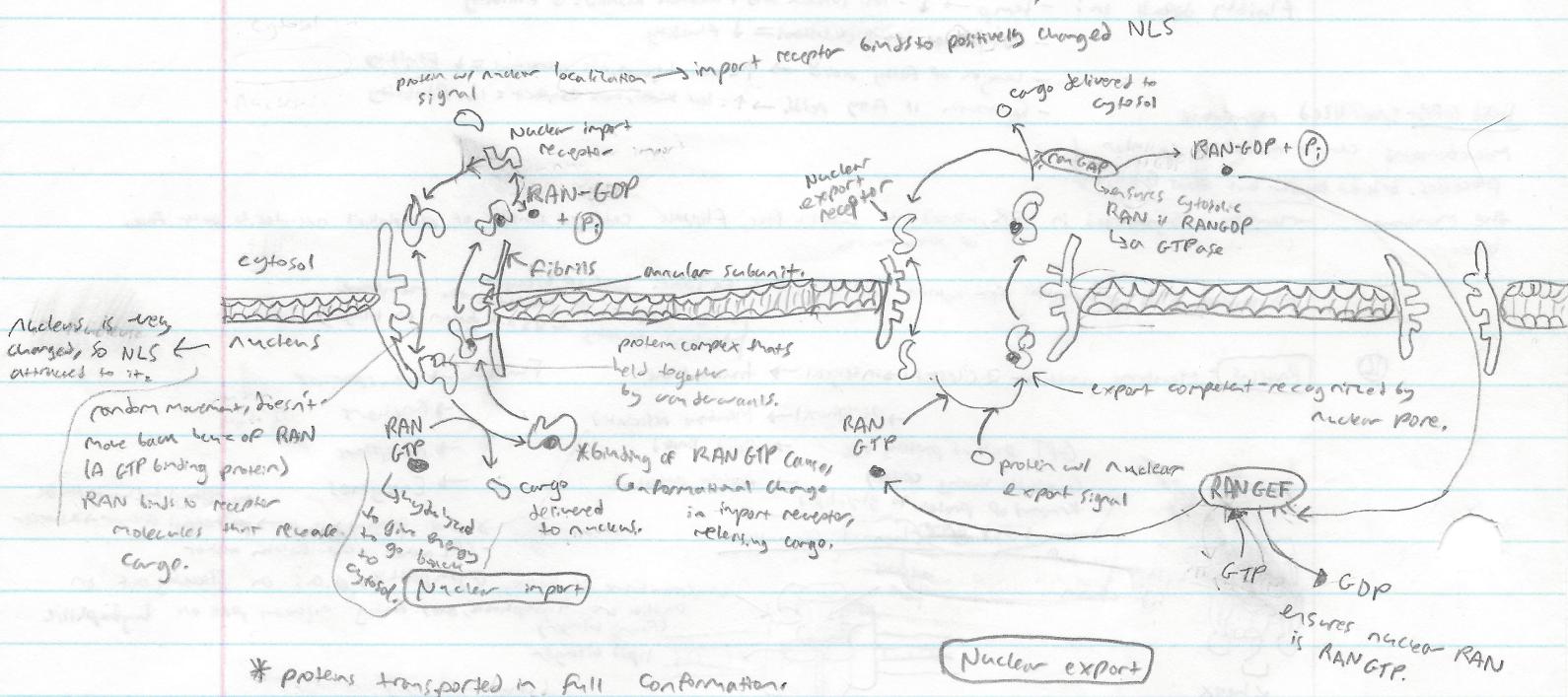
Nuclear pore complexes - bidirectional



- 300 distinct nuclear pores (specificity)

- 3000-4000 NPC per cell, ~125nm diameter, 500 molecules, 120nm, 100 diff proteins

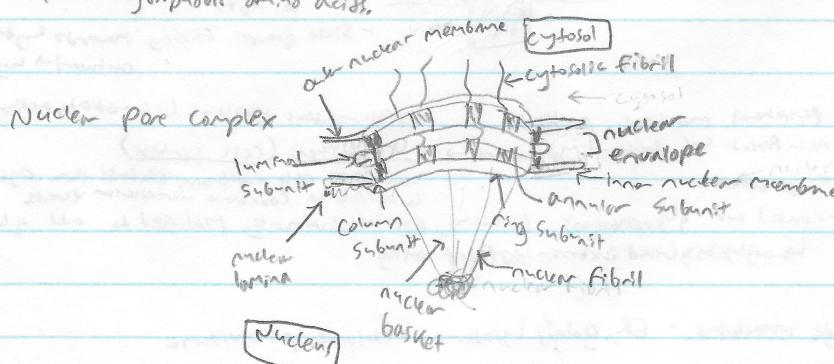
protein w/ nuclear localization signal  $\rightarrow$  import + receptor binds to positively charged NLS



\* Proteins transported in full conformations.

NES - nuclear export sequence. LX<sub>1-3</sub>LX<sub>2-3</sub>LXL X=any amino acid.

↳ rich in hydrophobic amino acids.

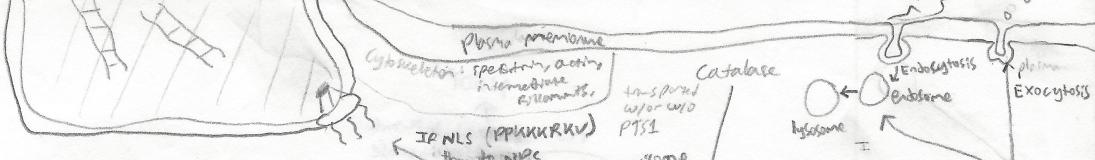


EF-tu  $\rightarrow$  brings charged tRNA with GTP to mRNA and ribosome complex in the A site

$\rightarrow$  If anticodon matches w/ codon, switch helix will release tRNA via hydrolysis of GTP  $\rightarrow$  GDP + Pi, latch opens

$\rightarrow$  EF-tu then leaves and gets phosphorylated and recycled, binding to a new charged tRNA.

$\rightarrow$  If anticodon does not match, GTP is not hydrolyzed and switch helix stays closed not releasing the tRNA.



### Protein Sorting and transport

